

GLYCOCINNASPERIMICIN D, A NEW MEMBER OF
GLYCOCINNAMOYLSPERMIDINE ANTIBIOTIC

KAZUYUKI DOBASHI, KATSUHIKO NAGAOKA, YOSHINORI WATANABE,
MAKOTO NISHIDA, MASA HAMADA, HIROSHI NAGANAWA,
TOMOHISA TAKITA, TOMIO TAKEUCHI
and HAMAO UMEZAWA

Institute of Microbial Chemistry
3-14-23 Kamiosaki, Shinagawa-ku, Tokyo 141, Japan

(Received for publication March 28, 1985)

Glycocinnasperimicin D was isolated from the fermentation filtrate of a strain of *Nocardia* sp. using various procedures of column chromatography. Glycocinnasperimicin D exhibited broad antibacterial spectrum. Its chemical structure was determined by NMR spectrometric analyses.

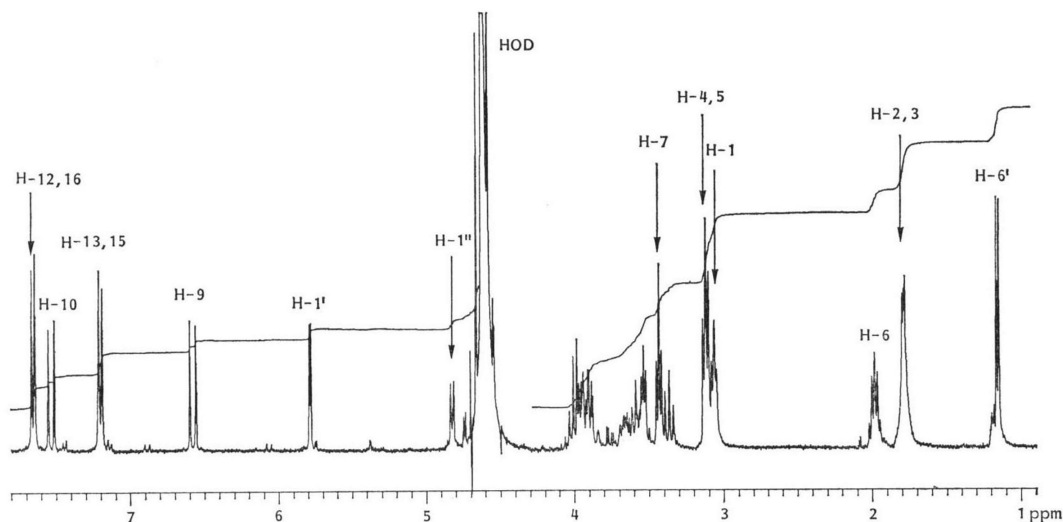
In the course of our screening for antibiotics, a new member of the glycocinnamoylspermidine group was found in the culture filtrate of *Nocardia* sp. strain MG615-7F6, and named glycocinnasperimicin D. This antibiotic has broad antibacterial spectrum. In this paper, we report on its fermentation, isolation, physico-chemical and biological properties. The strain also produced LL-BM123 β , γ_1 and γ_2^{11} .

Microorganism

The strain MG615-7F6 was isolated from a soil sample collected in Fuchu City, Tokyo, Japan. Cultural characteristics of the strain were related to those of BM123, NRRL 5646¹¹.

Fermentation and Isolation

A loopful of a mycelial suspension from a slant culture of strain MG615-7F6 was transferred into a seed medium containing glucose 1.0%, glycerol 1.0%, sucrose 1.0%, oat meal 0.5%, soy bean meal 2.0%, pressed yeast 1.0%, Casamino Acids 0.5%, CaCO₃ 1.0% (pH 7.0), and incubated with rotary shaking (180 rpm) for 3 days at 30°C. The seed culture (600 ml) was inoculated into 15 liters of a medium composed of glucose 1%, starch 1%, glycerol 1%, Polypeptone 0.5%, meat extract 0.5%, NaCl 0.5%, and CaCO₃ 0.32% (pH 7.4), in a 30-liter jar-fermentor. The fermentation was carried out at 28°C for 72 hours under an agitation of 350 rpm, aeration of 15 liters per minute. The cultured broth was adjusted to pH 5.0 with 1 M HCl and filtered. The filtrate (14.5 liters) thus obtained was passed through a column of Amberlite IRC-50 (Na⁺-, H⁺-form, 1: 1, 2.0 liters). After washing with water and 50% aqueous acetone, the adsorbed material was eluted with 1 M HCl-acetone (1: 1). The active fractions (assayed by a disk agar diffusion method using *Bacillus subtilis* PCI219) were collected and neutralized with 0.5 M NaOH. After evaporation of acetone, the solution was passed through a column of Amberlite XT-2 (80 ml) followed by elution with water. The active eluate was concentrated and lyophilized to give a pale yellow crude powder (124 mg). The powder was further purified by column chromatography on CM-Sephadex C-25 (150 ml) with linear gradient elution (NaCl, from 0.1 M to 1.2 M). The bioactive fraction was divided into two parts (fractions A and B);

Fig. 1. ^1H NMR spectrum of glycocinnasperimicin D (400 MHz, 40.5°C, D_2O).

the bioactive agent present in fractions A and B was adsorbed on a small column of CHP-20P (37 ~ 75 μm) (Mitsubishi Chemical Industries Limited) and desorbed with water. The activity in fraction A was separated further into two fractions. The early fraction of fraction A was glycocinnasperimicin D (7.1 mg) and the late one was identified as LL-BM123 γ complex (8.5 mg). The antibiotic from fraction B was identified as LL-BM123 β (10 mg) by direct comparison with authentic sample. These antibiotics could be distinguished by cellulose TLC: Eastman chromatogram sheet No. 6065, 1-propanol - pyridine - acetic acid - water (20: 10: 3: 12); glycocinnasperimicin D, Rf 0.28; LL-BM123 γ , Rf 0.24; LL-BM123 β , Rf 0.13. LL-BM123 α was not found in our culture broth.

Physico-chemical Properties and Structure

Glycocinnasperimicin D is obtained as a colorless powder, $[\alpha]_D^{25} +88^\circ$ (c 0.5, H_2O); mp $>300^\circ\text{C}$ (dec); it is a basic substance, easily soluble in water and very hygroscopic. It shows positive reaction to ninhydrin, Rydon-Smith, and Sakaguchi reagents. Elemental analysis did not give a satisfactory value; a characteristic of LL-BM123 antibiotics²⁾.

The secondary ion mass spectrum [SIMS, m/z 695 ($\text{M}+\text{H}^+$)] and the ^{13}C NMR spectrum indicated the molecular formula $\text{C}_{30}\text{H}_{50}\text{N}_{10}\text{O}_8$. The antibiotic showed UV maxima at 218 ($\log \epsilon$ 4.13) and 287 nm ($\log \epsilon$ 4.34) in water. In the ^1H NMR spectrum in D_2O (Fig. 1), two olefinic protons at δ 6.59 (1H, d, $J=16.0$ Hz), 7.53 (1H, d, $J=16.0$ Hz) and four aromatic protons at δ 7.20 (2H, d, $J=9.0$ Hz), 7.66 (2H, d, $J=9.0$ Hz) suggested the presence of a *p*-hydroxycinnamoyl moiety which is also present in LL-BM123 β and γ . The presence of a spermidine moiety was shown by methylene signals at δ 1.80 (4H, m), 1.99 (2H, m), 3.07 (2H, t), 3.13 (4H, t $\times 2$), and 3.43 (2H, t). Spin decoupling experiments indicated that the triplet signal at δ 3.43 was H-7 of spermidine; it is linked to the carbonyl group of cinnamic acid through an amide bond. Known glycocinnamoylspermidine antibiotics are composed of a cinnamoylspermidine aglycone and a trisaccharide moiety³⁾. However, glycocinnasperimicin D has a disaccharide moiety, as suggested by two anomeric protons at δ 5.79 (d, $J=3.3$ Hz) and 4.83 (d, $J=9.1$ Hz). A doublet signal at δ 1.06 (3H, d, $J=6.2$ Hz) and nine protons from δ 3.3 to 4.1 were attributed to the signals of methylenes and methines in the disaccharide moiety.

Table 1. ^{13}C NMR spectrum of glycocinnasperimicin D and LL-BM123 β , γ_1 (25 MHz, D_2O internal dioxane $\delta=67.4$).

Position	Glycocinnasperimicin D·HCl	LL-BM123 β ·H ₂ SO ₄ ^a	LL-BM123 γ_1 ·H ₂ SO ₄ ^a
1	37.1 (t) ^b		37.2
2	24.7 (t)		24.8
3	23.6 (t)		23.6
4	47.7 (t)		47.8
5	45.9 (t)		46.1
6	26.5 (t)		26.6
7	39.6 (t)		39.7
8	170.1 (s)		169.9
9	119.5 (d)		119.7
10	141.5 (d)		141.4
11	130.2 (s)		130.2
12, 16	130.6 (d)		130.6
13, 15	118.0 (d)		118.2
14	157.8 (s)	157.9 ^c	158.0 ^c
1'	96.0 (d)	96.0	96.4 (d)
2'	58.2 (d)	58.2	58.3 (d)
3'	69.2 (d)	69.2	69.2 (d)
4'	57.1 (d)	57.1	57.3 (d)
5'	70.8 (d)	70.8	71.0 (d)
6'	17.5 (q)	17.5	17.7 (q)
1''	82.5 (d)	82.2	82.4 (d)
2''	55.8 (d)	55.4	55.8 (d)
3''	75.4 (d)	73.2	73.4 (d)
4''	70.3 (d)	78.0	75.3 (d)
5''	67.2 (t)	64.6	64.6 (t)
1'''		101.8	98.2 (d)
2'''		53.2	56.1 (d)
3'''		57.0	60.8 (d)
4'''		66.9	70.0 (d)
5'''		66.7	66.9 (t)
2' C=N	158.2 (s)	158.3	158.3 (s)
4' C=O	159.8 (s)	159.7 ^c	159.7 (s) ^c
2'' C=O	162.1 (s)	161.7	162.1 (s) ^c
2''' C=O		162.1 ^c	160.8 (s)
			156.5 (s)

^a Taken from ref 3.^b Multiplicity in off-resonance spectrum.^c These assignments were referred to the result of ref 4.

The ^{13}C NMR spectrum of glycocinnasperimicin D showed six aromatic carbons, two olefinic carbons, and four carbonyl and/or imino carbons. The presence of two anomeric carbons at δ 96.0 and 82.5 and nine carbons in the disaccharide moiety was in accord with the result of the ^1H NMR analysis.

Comparison of ^{13}C NMR spectrum of glycocinnasperimicin D with those of LL-BM123 β and γ_1 (Table 1)^{3,4} indicated that the chemical shifts for the relevant carbons of these antibiotics, except for a substituent effect at C-4'', were very similar. The observed downfield shifts of C-3'', C-5'' and the upfield shift of C-4'' in glycocinnasperimicin D indicated a free hydroxyl group at C-4''. Therefore, the disaccharide moiety should correspond to that of LL-BM123 β without the distal sugar part.

Fig. 2. Structure of glycocinnasperimicin D.

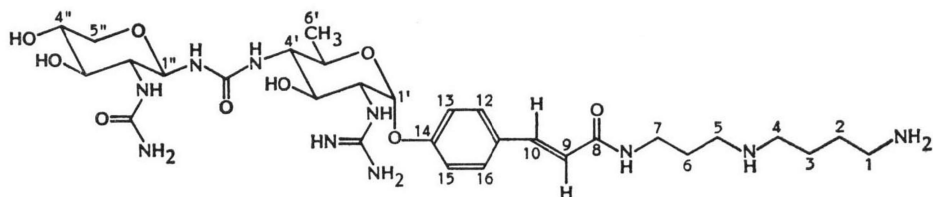


Fig. 3. IR spectrum of glycocinnasperimicin D (KBr).

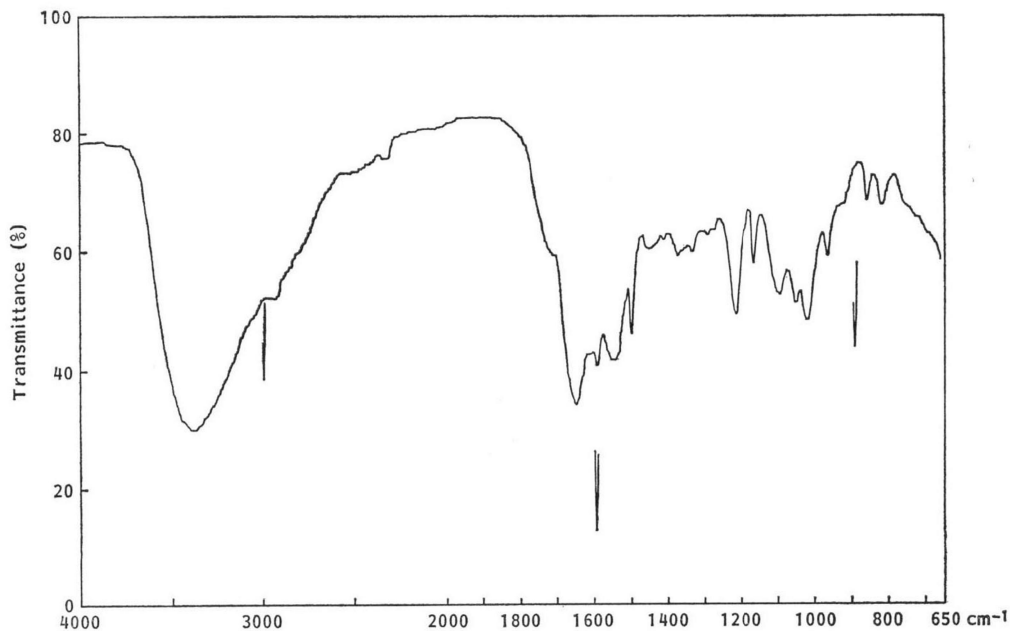


Table 2. Antimicrobial activity of glycocinnasperimicin D.

Organism	MIC ($\mu\text{g/ml}$)	Organism	MIC ($\mu\text{g/ml}$)
<i>Staphylococcus aureus</i> 209P	1.56	<i>Shigella dysenteriae</i> JS11910	1.56
<i>S. aureus</i> Smith	0.78	<i>S. flexneri</i> 4b JS11811	1.56
<i>S. aureus</i> Ap01	6.25	<i>S. sonnei</i> JS11746	3.12
<i>S. epidermidis</i> 109	3.12	<i>Salmonella typhi</i> T-63	0.39
<i>Corynebacterium bovis</i> 1810	3.12	<i>Proteus vulgaris</i> OX19	0.39
<i>Escherichia coli</i> NIHJ	0.39	<i>P. rettgeri</i> GN311	1.56
<i>E. coli</i> K-12	0.39	<i>P. rettgeri</i> GN466	0.78
<i>E. coli</i> JR66/W677	0.78	<i>Serratia marcescens</i>	6.25
<i>Mycobacterium smegmatis</i> ATCC 607	6.25	<i>Pseudomonas aeruginosa</i> A3	12.5
<i>Klebsiella pneumoniae</i> PC1602	1.56	<i>P. aeruginosa</i> GN-315	50

The C-1'' position linked to ureido nitrogen atom was reported by ELLESTAD *et al.*³⁾ to be an α -linkage in LL-BM123 β , γ_1 , and γ_2 . However, the signal of H-1'' was covered by water signals when ^1H NMR was taken at 23°C. The 400 MHz ^1H NMR spectra at 40°C (D_2O) of glycocinnasperimicin D and LL-BM123 β clearly showed the proton signal of H-1''; the doublet at δ 4.83 with J of 9.1 Hz,

δ 4.84 with J of 9.0 Hz, respectively. These data indicated that the linkage at C-1'' of both antibiotics should be β . From these results, we propose the structure of glycocinnasperimicin D as shown in Fig. 2.

Biological Effects

The antibacterial spectrum of glycocinnasperimicin D tested by serial agar dilution method is shown in Table 2. The 50% growth inhibitory concentration (IC_{50}) against Leukemia L-1210 cells was 2.0 μ g/ml.

LD_{100} (iv) of glycocinnasperimicin D in mice was 50 mg/kg and LD_0 (iv) 25 mg/kg.

Acknowledgment

We are grateful to Dr. D. B. BORDERS, American Cyanamid Company, for their gift of LL-BM123 β and γ_2 .

References

- 1) TRESNER, H. D.; J. H. KORSHALLA, A. A. FANTINI, J. D. KORSHALLA, J. P. KIRBY, J. J. GOODMAN, R. A. KELE, A. J. SHAY & D. B. BORDERS: Glycocinnamoylspermidines, a new class of antibiotics. I. Description and fermentation of the organism producing the LL-BM123 antibiotics. *J. Antibiotics* 31: 394~397, 1978
- 2) MARTIN, J. H.; M. P. KUNSTMANN, F. BARBATSCHI, M. HERTZ, G. A. ELLESTAD, M. DANN, G. S. REDIN, A. C. DORNBUSH & N. A. KUCK: Glycocinnamoylspermidines, a new class of antibiotics. II. Isolation, physicochemical and biological properties of LL-BM123 β , γ_1 and γ_2 . *J. Antibiotics* 31: 398~404, 1978
- 3) ELLESTAD, G. A.; D. B. COSULICH, R. W. BROSCARD, J. H. MARTIN, M. P. KUNSTMANN, G. O. MORTON, J. E. LANCASTER, W. FULMOR & F. M. LOVELL: Glycocinnamoylspermidines, a new class of antibiotics. 3. The structures of LL-BM123 β , γ_1 , and γ_2 . *J. Am. Chem. Soc.* 100: 2515~2524, 1978
- 4) TSOU, H.-R.; R. R. FIALA, P. C. MOWERY & M. W. BULLOCK: Biosynthesis of the spermidine and guanidino units in the glycocinnamoylspermidine antibiotic cinodine. *J. Antibiotics* 37: 1382~1387, 1984